## Fragilide E, a Novel Chlorinated 20-Acetoxybriarane from the Gorgonian Coral Junceella fragilis

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A novel chlorinated briarane, fragilide E (1), which possesses an unprecedented 20-acetoxy group, was isolated from the gorgonian coral *Junceella fragilis*. The structure of 1 was elucidated by spectroscopic methods.

In our research on new substances from octocorals collected in Taiwanese waters, a series of interesting briarane derivatives, including fragilides A–D,<sup>1–4</sup> have been isolated from *Junceella fragilis*. In this paper, we report the isolation, structure determination, and bioactivity of a novel briarane, fragilide E (1) (Chart 1), from further studies on *J. fragilis*. The structure of 1 was established by spectroscopic methods.

Specimens of *J. fragilis* (wet weight 780 g, dry weight 570 g) were collected by hand using scuba at the southern Taiwan coast in December 2002, at a depth of 20 m, and a voucher specimen was deposited in the NMMBA. The organisms were extracted with EtOH, and the residue was partitioned between EtOAc and H<sub>2</sub>O. The EtOAc layer was separated by silica gel column chromatography, using  $CH_2Cl_2$  and  $CH_2Cl_2$ -acetone mixtures of increasing polarity. Briarane **1** was eluted with  $CH_2Cl_2$ -acetone (8:1).

Fragilide E (1), 1.1 mg; mp 143–144 °C;  $[\alpha]_D^{25} + 13^\circ$  (*c* 0.05, CHCl<sub>3</sub>), was isolated as a white solid. The molecular formula of **1** was established as C<sub>28</sub>H<sub>37</sub>ClO<sub>13</sub> (10 degrees of unsaturation) from a sodiated molecule at *m*/*z* 639 in the ESIMS and was further supported by HRESIMS (*m*/*z* calcd: 639.1820; found: 639.1824,  $[C_{28}H_{37}^{35}ClO_{13} + Na]^+$ ). The IR spectrum of **1** showed bands at 3486, 1786, and 1742 cm<sup>-1</sup>, consistent with the presence of hydroxy,  $\gamma$ -lactone, and ester carbonyl groups. The <sup>13</sup>C NMR and DEPT spectra of **1** showed that this compound has 28 carbons, including six methyls, three sp<sup>3</sup> methylenes, an sp<sup>2</sup> methylene, nine sp<sup>3</sup> methines, three sp<sup>3</sup> quaternary carbons, and six sp<sup>2</sup> quaternary carbons. From the <sup>1</sup>H and

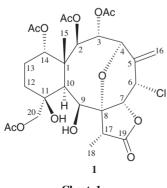


Chart 1.

Table 1. <sup>1</sup>H and <sup>13</sup>C NMR data and HMBC correlations for 1

<b>Table 1.</b> "H and "CNMR data and HMBC correlations for I			
C/H	$^{1}\mathrm{H}^{\mathrm{a}}/\delta$	$^{13}\mathrm{C}^{\mathrm{b}}/\delta$	HMBC $(H \rightarrow C)$
1		45.0 (s) <sup>d</sup>	
2	5.20 d (6.4) <sup>c</sup>	72.9 (d)	C-1, -3, -4, -10, -14, -15,
			acetate carbonyl
3	6.17 dd (10.8, 6.4)	65.5 (d)	C-1, -2, -4, -5, acetate carbonyl
4	4.36 d (10.8)	78.3 (d)	C-2, -3, -5, -6, -8, -16
5		135.2 (s)	
6	5.58 d (2.8)	54.8 (d)	C-4
7	4.74 d (2.8)	80.3 (d)	C-5, -6, -8, -9
8		82.9 (s)	
9	4.75 d (3.2)	76.5 (d)	C-1, -11, -17
10	2.18 d (2.0)	43.1 (d)	C-1, -8, -9, -11, -15
11		75.8 (s)	
12/12'	1.76 m; 1.69 m	30.4 (t)	C-10, -11, -14
13	1.73 m (2H)	20.7 (t)	C-1, -11, -14
14	5.02 br s	74.3 (d)	C-2, -12
15	1.57 s	17.5 (q)	C-1, -2, -10, -14
16a	5.29 d (2.0)	118.6 (t)	C-4, -5, -6
b	5.54 d (2.0)		C-4, -5, -6
17	2.59 q (7.2)	49.7 (d)	C-9, -18, -19
18	1.31 d (7.2)	8.3 (q)	C-8, -17, -19
19		175.3 (s)	
20a	3.98 d (11.2)	72.3 (t)	C-10, -12, acetate carbonyl
b	4.24 d (11.2)		C-10, -11, -12, acetate carbonyl
OH-9	2.88 d (3.2)		C-8, -9
OH-11	2.56 d (2.0)		C-10, -11
2-OAc		170.3 (s)	
	2.03 s	20.5 (q)	Acetate carbonyl
3-OAc		169.3 (s)	
	1.97 s	20.5 (q)	Acetate carbonyl
14-OAc		170.1 (s)	
	2.01 s	20.9 (q)	Acetate carbonyl
20-OAc		171.5 (s)	
	2.15 s	20.8 (q)	Acetate carbonyl

Spectra recorded at <sup>a</sup>400 and <sup>b</sup>100 MHz in CDCl<sub>3</sub> at 25 °C, respectivley. <sup>c</sup>J values (in Hz) in parentheses. <sup>d</sup>Multiplicity deduced by DEPT and indicated by usual symbols.

<sup>13</sup>C NMR spectra (Table 1), briarane 1 was found to possess four acetoxy groups ( $\delta_{\rm H}$  2.15, 2.03, 2.01, 1.97, each 3H × s;  $\delta_{\rm C}$  171.5, 170.3, 170.1, 169.3, each s), a  $\gamma$ -lactone moiety ( $\delta$  175.3, s), and an exocyclic carbon–carbon double bond ( $\delta_{\rm C}$  135.2, s; 118.6, t;  $\delta_{\rm H}$  5.54, 1H, d, J = 2.0 Hz; 5.29, 1H, d, J = 2.0 Hz). Thus, from the NMR data, six degrees of unsaturation were accounted for, and 1 was identified as a tetracyclic compound.

The gross structure of **1** was determined using 2D NMR studies. From the  ${}^{1}H{-}^{1}HCOSY$  spectrum of **1**, five different structural units were identified (Figure 1), which were assembled with the assistance of an HMBC experiments (Table 1 and Figure 1). The HMBC correlations between protons and quater-

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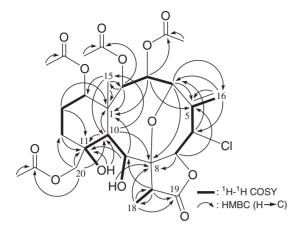


Figure 1. The <sup>1</sup>H–<sup>1</sup>HCOSY and selective key HMBC correlations of 1.

nary carbons of 1, such as H-2, H-3, H-9, H-10, H<sub>3</sub>-15/C-1; H-3, H-4, H-7, H<sub>2</sub>-16/C-5; H-4, H-7, H-10, H<sub>3</sub>-18, OH-9/C-8; H-9, H-10, H<sub>2</sub>-12, H<sub>2</sub>-13, H-20b/C-11; and H-17, H<sub>3</sub>-18/C-19, permitted elucidation of the carbon skeleton. An exocyclic double bond attached at C-5 was established by the HMBC correlations between H<sub>2</sub>-16/C-4, -5, -6 and H-4/C-16; and further confirmed by allylic couplings between H<sub>2</sub>-16 and H-6. The ring junction C-15 methyl group was positioned at C-1 from the HMBC correlations between H<sub>3</sub>-15/C-1, -2, -10, -14; H-2/C-15; and H-10/ C-15. The presence of acetate esters positioned at C-2, C-3, and C-20 were established by the HMBC correlations between protons H-2 (\$ 5.20), H-3 (\$ 6.17), H-20a/b (\$ 3.98, 4.24) and acetate carbonyls ( $\delta$  170.3, 169.3, 171.5). The remaining acetoxy group was positioned at C-14 as indicated by analysis of <sup>1</sup>H-<sup>1</sup>HCOSY correlations and characteristic NMR signals analysis, although no HMBC correlation was observed between H-14 and acetate carbonyl. The hydroxy proton signal appearing at  $\delta$  2.88 (1H, d, J = 3.2 Hz) was revealed by its <sup>1</sup>H-<sup>1</sup>HCOSY and HMBC correlations to H-9 ( $\delta$  4.75, 1H, d, J = 3.2 Hz) and C-9 ( $\delta$  76.5, d), respectively, indicating its attachment to C-9. The presence of an 11-hydroxy group was deduced from the HMBC correlations between a hydroxy proton ( $\delta$  2.56) with an oxygenated quaternary carbon resonating at  $\delta$  75.8 (C-11) and C-10 methine ( $\delta$  43.1). This observation was also evidenced by the HMBC correlations between one proton of acetoxymethyl ( $\delta$  4.24, H-20b) and C-11 oxygenated quaternary carbon and C-12 methylene.

The intensity of sodiated molecule  $(M + 2 + Na)^+$  isotope peak observed in ESIMS  $[(M + Na)^+:(M + Na + 2)^+ = 3:1]$ was strong evidence of the presence of a chlorine atom in **1**. The methine unit at  $\delta$  54.8 (d) was more shielded than that expected for an oxygenated C-atom, and was correlated to the methine proton appearing at  $\delta$  5.58 in the HMQC spectrum. The latter methine signal was <sup>3</sup>*J*-correlated with C-4 ( $\delta$  78.3, d), proving the attachment of a chlorine atom at C-6. Furthermore, an HMBC correlation between H-4 ( $\delta$  4.36) and an oxygenated quaternary carbon appearing at  $\delta$  82.9 (s, C-8) suggested the presence of C-4/8 ether linkage. These data, together with the HMBC correlations between H-17/C-9, -18, -19; and H<sub>3</sub>-18/C-8, -17, -19, unambiguously established the molecular framework of **1**.

The relative stereochemistry of **1** was elucidated by analysis of NOESY correlations (Figure 2) and by coupling constant

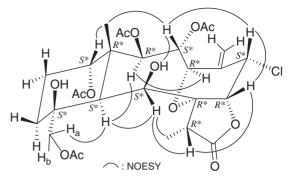


Figure 2. Selective NOESY correlations of 1.

analysis. The NOESY correlations between H-10 and H-2, H-9, and one proton of C-20 methylene ( $\delta$  3.98, H-20a) indicated that these protons were situated on the same face; they were assigned as  $\alpha$  protons, as C-15 methyl was  $\beta$ -oriented and H<sub>3</sub>-15 did not show correlation with H-10. H-10 exhibited a small coupling constant with OH-11 by a long-range w-coupling (J = 2.0 Hz), indicating the 11-hydroxy group was  $\beta$ -oriented. H-14 was found to exhibit a response with H<sub>3</sub>-15 but not with H-10, revealing the  $\beta$ -orientation of this proton. The correlations observed between H-3/H<sub>3</sub>-15, H-3/OH-9, H-3/H-6, H-6/H-7, and H-7/H-17, reflected the  $\beta$ -orientation of protons attached at C-3, C-6, C-7, and C-17. Furthermore, H-4 was found to show a correlation with H-2; and a large coupling constant was found between H-4 and H-3 (J = 10.8 Hz), indicating the dihedral angle between H-4 and H-3 is approximately 180° and H-4 has an  $\alpha$ -orientation at C-4. Based on the above findings, the chiral centers of 1 were assigned as 1R\*, 2R\*, 3S\*, 4R\*, 6S\*, 7R\*, 8R\*, 9S\*, 10S\*, 11S\*, 14S\*, and 17R\*.

It is worth noting that briarane metabolites possessing an oxygenated C-20 group, such as acetoxy,<sup>5</sup> hydroxy,<sup>6</sup> or carboxylic acid groups,<sup>7</sup> are rarely found. The structure of **1** was found to be similar with that of the first briarane containing a carboxylic acid group, juncin N,<sup>7</sup> however, fragilide E (**1**) is the second 20-acetoxybriarane ever discovered.<sup>5</sup> In biological activity experiment, **1** displayed 16.6% inhibitory effect on superoxide anion generation and 17.7% inhibitory effect on elastase release by human neutrophil at 10 µg/mL, respectively.

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## References

- P.-J. Sung, M.-R. Lin, W.-C. Chen, L.-S. Fang, C.-K. Lu, J.-H. Sheu, Bull. Chem. Soc. Jpn. 2004, 77, 1229.
- 2 P.-J. Sung, Y.-P. Chen, Y.-M. Su, T.-L. Hwang, W.-P. Hu, T.-Y. Fan, W.-H. Wang, Bull. Chem. Soc. Jpn. 2007, 80, 1205.
- 3 P.-J. Sung, M.-R. Lin, Y.-D. Su, M. Y. Chiang, W.-P. Hu, J.-H. Su, M.-C. Cheng, T.-L. Hwang, J.-H. Sheu, *Tetrahedron* **2008**, *64*, 2596.
- 4 P.-J. Sung, C.-H. Pai, Y.-D. Su, T.-L. Hwang, F.-W. Kuo, T.-Y. Fan, J.-J. Li, *Tetrahedron* **2008**, *64*, 4224.
- 5 C. Tanaka, Y. Yamamoto, M. Otsuka, J. Tanaka, T. Ichiba, G. Marriott, R. Rachmat, T. Higa, J. Nat. Prod. 2004, 67, 1368.
- 6 J.-H. Su, P.-J. Sung, Y.-H. Kuo, C.-H. Hsu, J.-H. Sheu, *Tetrahedron* 2007, 63, 8282.
- 7 P.-J. Sung, T.-Y. Fan, L.-S. Fang, J.-H. Sheu, S.-L. Wu, G.-H. Wang, M.-R. Lin, *Heterocycles* 2003, 61, 587.